

# Correlation between the autonomic nervous system and neoplastic disease

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**Summary.** The physiological role of the autonomic nervous system (ANS) includes maintenance of homeostasis and response to stressors. Sympathetic nervous system (SNS) is involved at early stages of tumorigenesis via  $\beta$ -adrenergic signaling and via central and local norepinephrine/epinephrine release from SNS nerve fibers. Parasympathetic nervous system (PNS) promotes invasion and dissemination of cancers using cholinergic receptors. In this paper, we review published evidence regarding the impact of SNS and PNS on solid tumor development, and discuss the importance of those findings for clinic and prevention.

**Key words:** autonomic nervous system, sympathetic nervous system, parasympathetic nervous system, neoplasm

## Physiological function of the autonomic nervous system

Autonomic nervous system (ANS) consists of two main parts: sympathetic nervous system (SNS, noradrenergic) and parasympathetic nervous system (PNS, cholinergic). Both of them play a significant role in maintaining homeostasis and stress response. Physiological mechanisms involved in the autonomic activity include neural outflow, synthesis, release and degradation of transmitters, ganglionic regulation and receptor-mediated effects.

Sympathetic fibers participate in many physiological processes, inter alia regulating efferent sympathetic nerve outflow and generating its differential patterns, synchronizing neural activity of various target tissues and organs and thus, controlling function thereof. Moreover, acting via  $\beta$ -adrenergic signaling pathway, SNS induces 'fight-or-flight' stress response. SNS fibers innervate all major organs, releasing catecholamine neurotransmitters (norepinephrine and epinephrine) in response to a physiological threat to homeostasis or acute sympathetic activation. Norepinephrine and epinephrine, the me-

tabolites of amino acid, tyrosine, are released from the adrenal medulla during a stress reaction. Their biological effects include an increase in blood pressure, stimulation of liver glycogenolysis and airway dilation.

Parasympathetic nervous system provides innervation to many organs, controlling a number of vital physiological functions, such as heart rate, endocrine activity, digestion, gastrointestinal motility, inflammation and immune response. Vagus nerve, the X cranial nerve, and its branches contain 80% of afferent sensory fibers and 20% of efferent motor fibers, and form a communicating sensory pathway between the central nervous system and peripheral tissues. The main parasympathetic neurotransmitter is acetylcholine, acting via five types of muscarinic receptors (chrm1 – chrm5) and two types of nicotinic receptors (muscle-type, N1, and neuronal-type, N2).

## Autonomic contribution to neoplastic processes – Underlying mechanisms

Tumor progression and dissemination depend on intrinsic prosperities of cancer cells, such as self-

renewal and the ability to migrate (invasiveness). Tumor stroma, containing fibroblasts, endothelial cells and immune cells, may interact with cancer cells (1, 2). This microenvironment regulates growth of a primary tumor and formation of metastases. Both stromal cells of the tumor and signaling molecules interact directly and indirectly with the nervous system (3). The metastatic cascade involves two main steps. During the first stage, tumor cells migrate from the primary tumor to the site of hematogenous and lymphogenous dissemination. During the second stage, the cells extravasate from the circulation and invade surrounding tissues (4, 5).

Both sympathetic and parasympathetic components of the autonomic nervous system play vital roles during the development and spread of solid tumors, albeit at different stages of tumorigenesis. Likewise in leukocyte and fibroblast migration, neurotransmitters regulate also the migratory activity of cancer cells. Cancer cells may migrate along nerve fibers; this phenomenon, referred to as perineural invasion, is associated with poorer prognosis (6-10). SNS mediates tumor initiation and progression through a variety mechanisms.  $\beta$ -adrenergic signaling promotes the inhibition of DNA damage repair and p53-associated apoptosis via several molecular pathways (e.g.  $\beta$ -arrestin-induced activation of the AKT signaling pathway); as a result, SNS may contribute to tumor initiation and/or chromosomal instability (1, 2). Macrophages play an important role, modulating tumor microenvironment and promoting metastasis.  $\beta$ -adrenergic signaling stimulates recruitment of macrophages to tumor parenchyma via chemotactic factors, such as macrophage colony stimulating factor (CSF1). Moreover,  $\beta$ -adrenergic signaling contributes to an increase in tumor-associated macrophage density, promoting myelopoietic development of monocyte precursors in the spleen and bone marrow. Finally,  $\beta$ -adrenergic signaling stimulates macrophage expression of gene programs that initiate tumor progression within the tumor microenvironment.

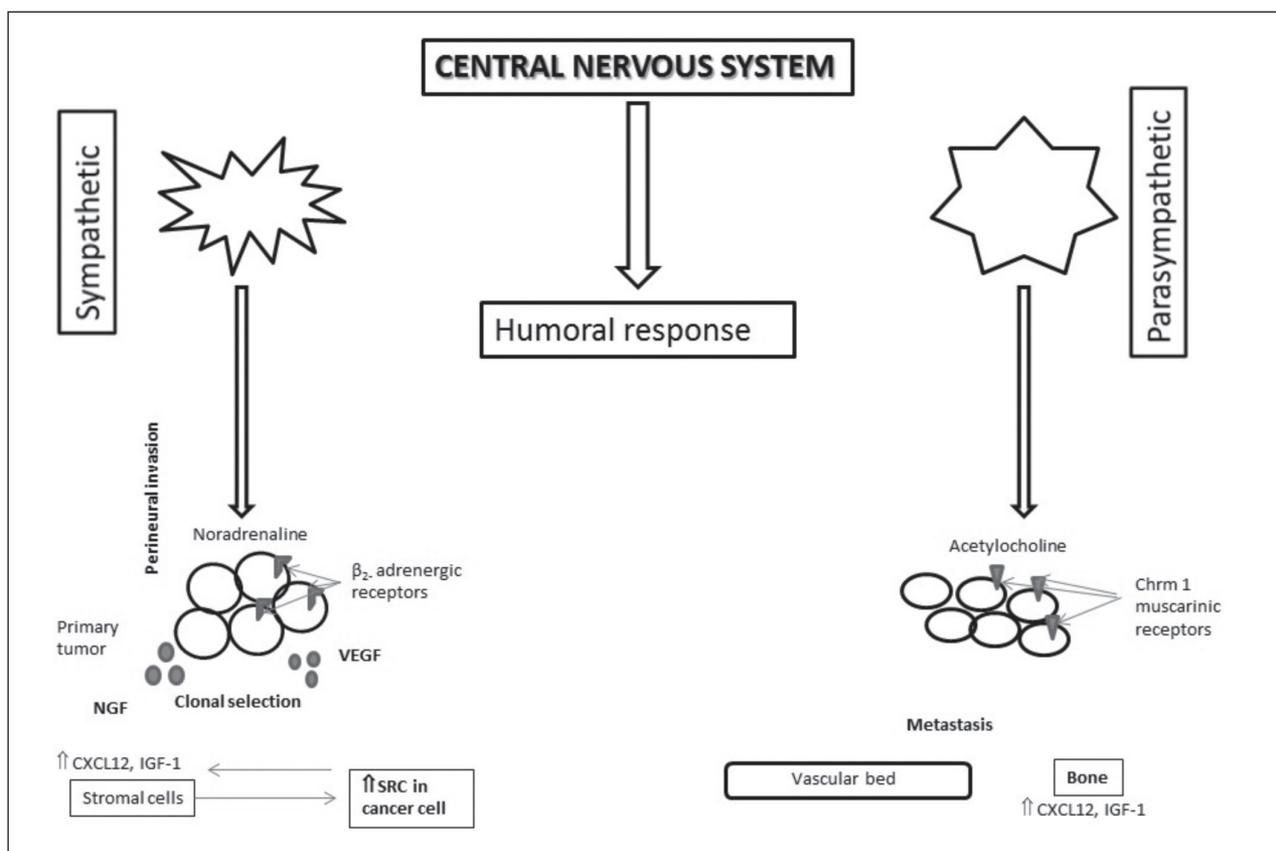
Additionally,  $\beta$ -adrenergic signaling may also modulate various growth and survival pathways, among them programmed cell death mediated by focal adhesion kinase (FAK) (11). SNS generally promotes tumor progression through an array of pleiotropic mo-

lecular alterations in the microenvironment of primary tumor (12). Tumor cells may spread via lymphatic and blood vessels, and along serous membranes (13). Perineural invasion results from interactions between cancer cells and nerve fiber microenvironment. Sympathetic nerves may contribute to the perineural invasion and stimulate tumor growth. Cancer cells stimulate the expression of stromal cell-derived factor 1 (also referred to as C-X-C motif chemokine ligand 12, CXCL12) and insulin-like growth factor 1 (IGF-1) in stromal cells; this eventually contributes to the selective growth of cancer cells clones that are hypersensitive to these factors and primed for spread in CXCL12- and IGF-1-enriched bone marrow (14) (Fig. 1). This pathway is also implicated in spread of cancer cells in patients who do not show the evidence of blood and/or lymphatic metastases (15, 16). A reciprocal signaling interaction between tumor cells and nerves contributes to peripheral nerve invasion. Neurotrophic factors (NGF) and axonal guidance molecules are pivotal for axonal growth (17, 18). However, these molecules and their receptors are also localized in tumor cells and hence, the latter can bind to the neurites (19, 20). According to Liebig et al., peripheral nerve invasion can be diagnosed whenever tumor cells are present within any of the three layers of the nerve sheath (epineurium, perineurium, endoneurium), or if tumor foci exist outside the nerve, involving at least 33% of its circumference (16). Also ANS may interfere with angiogenesis and modulate tumor microenvironment. Cancer cells can release neurotropic factors, such as axon guidance molecules, which stimulate the growth of nerve fibers, blood vessels and lymphatics (neovascularization and lymphangiogenesis, respectively) supplying the tumor (9, 21, 22).

Moreover, neoplastic cells can release angiogenic factors, such as vascular endothelial growth factor (VEGF), and angiogenic chemokines, which also promote neovascularization (21, 23). Tumors with diameters greater than 1 cm cannot be effectively supplied with nutrients without the development of new blood vessels (neovascularization) (21). NGF, acting alone or in combination with 6-hydroxy-dopamine (6-OHDA), was shown to stimulate neovascularization in the superior cervical ganglia of newborn rats; similar effect was also observed in breast and prostate cancers, whereby

NGF initiated neoangiogenesis and apoptosis, and modulated the severity cancer-induced bone pain (24-26). VEGF is a mitogen for endothelial cells, and promotes angiogenesis in vivo. Both VEGF and its receptors (expressed on sympathetic nerve fibers innervating arteries) promote vascular sympathetic innervation. VEGF promotes sympathetic axon growth. Its effects on vascular sympathetic innervation are modulated by other vascular-derived neuronal growth factors. Inhibition of VEGF was shown to prevent reinnervation in vivo (27). Aside from angiogenesis, VEGF was also shown to modulate lymphangiogenesis; since it shows a neurotrophic activity within the peripheral nervous system, this factor may also contribute to neurogenesis (23, 28-30).

Survival and growth of cancer cells may be also promoted by nerve-derived growth factors released from nerve fibers located in close vicinity of the tumor (31, 32) (Fig. 1). Fibroblast growth factor (FGF) regulates synaptogenesis and neuronal migration and therefore, may also contribute to the development of tumor innervation (33). Aside from the stimulation of hematopoiesis, granulocyte-colony stimulating factor (G-CSF), a hematopoietic stem cell mobilizer with established clinical application, may also modulate neuronal activity. This was confirmed in a mouse study, in which G-CSF affected survival of autonomic nerve fibers, which resulted in growth and dissemination of prostate cancer cells (34). Secretion of soluble factors from prostate cancer cells may contribute to nerve



**Figure 1.** The role of sympathetic and parasympathetic regulation in cancer progression and dissemination. Sympathetic nerves contribute to perineural invasion of cancer cells, and stimulate tumor growth. Cancer cells induce the expression of CXCL12 and IGF-1 in stromal cells, which promotes selective growth of clones that are hypersensitive to these factors and primed for metastasis in CXCL12- and IGF-1-enriched bone marrow. In turn, parasympathetic nerves play a role in cancer cell expansion and dissemination (according to del Toro et al., modified) (14).

sprouting and/or branching (10). Due to presence of neuro-neoplastic synapses and expression of receptors for neural markers, cancer cells may effectively communicate with neurons. Neurotransmitters reach cancer cells via neuro-neoplastic synapses, stimulating their migration and therefore, contributing to tumor spread (35). Neuro-neoplastic synapses are functional units, rather than morphological entities (8). Neurotransmitters may stimulate the release of acetylcholine from a subset of CD4(+) T cells and thus, indirectly activate other immune cells, for example via upregulation of  $\alpha 7$ -nicotinic acetylcholine receptors on macrophages (36).

### **Autonomic nervous system and immunological processes**

Both sympathetic and parasympathetic nervous system participate in neuroimmune processes. Immune cells (especially lymphocytes) express adrenergic and muscarinic receptors, as well as the receptors for acetylcholine (ACh), choline acetyltransferase (ChAT) and acetylcholinesterase (AChE). Neurotransmitters released from the sympathetic and parasympathetic nerve endings bind to their receptors and initiate immunomodulatory response. The interactions between SNS and immune system involve signaling mechanisms that provide a link between neurotransmitters and neuromodulators, co-transmitters (ATP and adenosine), adrenergic receptors and, albeit not necessarily, immune cells, cytokines and bacteria. SNS innervation of lymphoid tissue (the spleen and lymph nodes) modulates the evolution of peripheral immune response via cytokines, and promotes humoral immune responses at an expense of cellular immunity (12). Neuronal and neuroendocrine pathways are involved in communication between the nervous and immune system. Vagus nerve has been implicated as a component of the neural pathway transmitting signals from the peripheral immune system to the brain. Also cutaneous sensory afferents have been postulated to provide a communication pathway to central neural circuits. The immune to CNS communication is mediated by three non-neuronal mechanisms: cytokine transport system, brain structures containing blood vessels with fenestrated capillaries, and some mol-

ecules (e.g. prostaglandins, cytokines) that may reach the brain parenchyma (37). Hence, the neuronal system may contribute to cancer spread via the immune system (38). Chronic stress and depression may predispose to tumor spread due to impairment of immune response associated with decreased number of cytotoxic T-cells and natural killer (NK) cells (39).

Cholinergic anti-inflammatory pathway may involve cholinergic-adrenergic interactions at peripheral sites, autonomic ganglia and lymphoid targets (37, 40-43). The synthesis of immune and inflammatory mediators (cytokines, chemokines and free radicals) and the activity of various lymphoid cells are modulated due to activation of adrenergic receptors (37, 40-41). The activation of adrenergic receptors may affect synthesis of many compounds, among them tumor necrosis factor-alpha (TNF $\alpha$ ), interleukins (IL-6, IL-10 and IL-12), chemokine macrophage inflammatory protein 1 alpha and a free radical, nitric oxide (41).

Analysis of genomic profiles identified specific patterns related to various disease entities. For example, one previous study revealed commonalities in a genetic signature, which turned out to be associated with both  $\beta$ -adrenergic receptor and cancer (3). Breast cancer and  $\beta$ -adrenergic pathway were demonstrated to share some genetic signatures, namely IL-6, MMP9, MMPI, FOSB, LCK, ERG, CCL2I, RHOJ, IGFI and ETSI; this observation supports the hypothesis on a link between  $\beta_2$ -adrenergic receptor and breast cancer pathway, and may constitute a foundation for new anticancer therapies based on adrenergic receptor strategies (44).  $\beta$ -adrenergic pathway may influence oncogene pathways, such as *Her2* and SRC (45-46). Indeed,  $\beta$ -adrenergic pathway has already been shown to stimulate the phosphorylation of SRC by protein kinase A, which resulted in SRC-mediated activation of a complex phosphoproteomic network promoting tumor growth and invasion in vivo (45). Furthermore, a positive correlation between  $\beta_2$ -AR level ( $\beta_2$ -adrenergic receptor) and *Her2* status was reported in breast cancer cells. Activation of  $\beta$ -adrenergic receptor by a catecholamine contributed to the upregulation of *Her2* mRNA expression and to the stimulation of signal transducer and activator of transcription 3 (STAT3); the latter molecule activates ERBB promoter to stimulate gene transcription (47). Furthermore,  $\beta$ -adrenergic signaling

pathway may prevent DNA repair and suppress p53 level and t53-associated apoptosis via molecular pathways, such as Rad3-related (ART)/p21 pathway (1-2, 48-49). While this mechanism was already demonstrated to be sufficient for an increase in the number of spontaneous chromosomal aberrations in the tissues, we still do not know whether the  $\beta$ -adrenergic inhibition of DNA repair may contribute to the initiation of spontaneous tumorigenesis *in vivo* (12).

### Sympathetic Nervous System (SNS)

The effects of catecholamine neurotransmitters are mediated by receptors from  $\alpha_1$ ,  $\alpha_2$ ,  $\beta_1$ ,  $\beta_2$  and  $\beta_3$  families.  $\beta$ -adrenergic receptors are G-protein coupled receptors that activate adenylate cyclase to “synthesize” intracellular 3',5'-cyclic adenosine monophosphate (cAMP); the latter activates protein kinase A (PKA) to phosphorylate serine or threonine in target proteins. PKA is involved in regulation of cellular metabolism, growth, differentiation, secretion, motility, neurotransmission and gene transcription. The second cAMP effector is guanine nucleotide Exchange Protein by Adenylate Cyclase (EPAC). EPAC signaling may alter cellular morphology, motility and secretion (3).  $\beta$ -adrenergic signaling pathway regulates activity of various cells, among them epithelial cells, vascular myocytes, myeloid and lymphoid immune cells (50). All these cells express adrenergic receptors and therefore, are prone to the autonomic control.  $\beta$ -adrenergic signaling contributes to migration of cancer cells (51-53) and regulates VEGF-dependent angiogenesis (54-56) (Fig. 1). Also matrix metalloproteinase-related enhanced tissue invasion remains under the  $\beta$ -adrenergic control (56-58). Additionally, proapoptotic protein BAD (B-cell lymphoma 2-associated death promoter) may contribute to the development of chemotherapy resistance, acting via  $\beta$ -adrenergic receptor (11).  $\beta$ -adrenergic receptors, primarily  $\beta_2$ , are expressed at metastatic sites: in the brain, lungs, liver, lymphoid tissue, etc. (54, 59-60).  $\beta$ -adrenergic signaling pathway modulates the migratory potential and invasiveness of cancer cells via alterations in tumor gene expression and via upregulation of matrix metalloproteinases (MMP-6 and MMP-9) (57). Moreover, adrenergic

receptors may modulate pro-metastatic consequences of tumor immune response. Activation of adrenergic receptors initiates infiltration of tumor tissues by macrophages and promotes a pro-metastatic gene expression signature. A consequence of macrophage infiltration is overexpression of macrophage-derived factors (e.g. COX2, MMP-9, VEGF) (61-62). Due to the activation of  $\beta_2$ -adrenoreceptor-cAMP-protein kinase A pathway, epinephrine and norepinephrine inhibit secretion of type 1 proinflammatory cytokines: interleukin-12 (IL-12), TNF $\alpha$  and interferon- $\gamma$  by antigen-presenting cells and Th1 helper cells. Furthermore, these endogenous catecholamines promote secretion of type 2 anti-inflammatory cytokines: interleukin-10 (IL-10) and transforming growth factor  $\beta$  (TGF $\beta$ ) (40). These neurotransmitters can suppress Th1 response, which results in the inhibition of cell-mediated immunity and predominance of humoral immune response. Activation of SNS during the course of immune response may limit the inflammatory reaction, promoting accumulation of neutrophils and stimulating more specific humoral response; this protects the host against unfavorable effects of proinflammatory cytokines and other compounds released from activated macrophages (40). SNS is involved at early stages of tumorigenesis via  $\beta$ -adrenergic signaling, and systemic and local release of norepinephrine/epinephrine from sympathetic nerve fibers (3, 6, 9, 63-65). This results in an increase the number of axons and promotes ramification thereof. Furthermore, cancer cells can produce axon guidance molecules and neurotrophic factors, such as NGF (66-67). The axon guidance molecules contribute to the development of new blood vessels. Four families of the guidance molecules exist: netrins, slits, ephrins and semaphorins. Netrins are bound by UNC5 and deleted in colorectal cancer (DCC) receptors, whereas slits, ephrins and semaphorins interact with slits-roundabout receptors (Robos), Ephrin receptors (Eph) and plexins/neuropilins, respectively (68). While netrins are known to prevent cancer cell apoptosis and thus, participate in tumorigenesis, their role in cancer cell migration is yet to be established (69). Semaphorins play an important regulatory role in carcinogenesis (70). For example, SEMA3B and SEMA3F act as tumor suppressors (71), and overexpression of SEMA3B was demon-

strated to cause apoptosis in breast and lung cancer cell lines (72). Ephrins and their receptors are involved in carcinogenesis through diverse mechanisms; for example, EphA4 receptor was shown to induce proliferation and migration of glioblastoma cells (73). Moreover, both ephrins and their receptors are the regulators of tumor microenvironment, providing a link between cancer cells and surrounding stroma (74).

In turn, vascular growth factors, e.g. VEGF, were shown to control the development of sympathetic innervation (75). Sympathetic axons follow the newly developed arteries that release neurotrophic factors, such as endothelin and artemin (76-77). However, this relationship seems to be bidirectional, since also sympathetic nervous system was demonstrated to provide some regulatory input to angiogenesis and arteriogenesis (78-79). These complex interactions explain tumor's potential to develop its own neurovascular network. SNS may influence the neoplastic growth, modulating tumor microenvironment in both primary and metastatic target sites, via  $\beta$ -adrenergic regulation of myelopoiesis (80-82).  $\beta$ -adrenergic pathway influences expression of genes involved in oncogenesis and tumor spread, inter alia those controlling inflammatory processes, neoangiogenesis, cellular immune response and programmed cell death (12, 83). Furthermore,  $\beta$ -adrenergic signaling pathway promotes development of hematological malignancies, interfering with stem cell biology and physiological hematopoiesis (12).

Stress is associated with the release of IL-6, a proinflammatory cytokine playing pivotal role in cancer spread. Stress stimulates the sympathetic-adrenal-medullary axis to release catecholamine hormones, such as norepinephrine (9). The latter may be involved in upregulation of metalloproteinases (MMP-2, MMP-9) and VEGF; due to their modulatory effects, the proinflammatory cytokines may inter alia contribute to greater invasiveness of nasopharyngeal carcinomas (56). Moreover, stress induces norepinephrine and  $\beta$ -adrenergic receptors under IL-6 regulation (56, 84). Aside from being a key inflammatory mediator, transcription factor NF- $\kappa$ B may also contribute to carcinogenesis and tumor spread. One direct transcriptional target for NF- $\kappa$ B is neuronal guidance molecule, netrin-1, which was shown to be upregulated during the course of inflammatory processes.

$\beta$ -adrenergic receptors were found in the breast, prostate and pancreatic cancer cells as well as in melanoma cells; their activation by catecholamine neurotransmitters resulted in enhanced activity of those cells (9, 85-88).

### **Parasympathetic Nervous System (PNS)**

Parasympathetic nervous system promotes invasion and dissemination of cancer cells, acting via cholinergic receptor muscarinic 1 (chrm 1) expressed in tumor stroma (6, 7, 63, 89). The activation of chrm1 by acetylcholine results in cancer spread. This phenomenon has been first demonstrated in a mouse cancer model, and then confirmed in patients with prostatic adenocarcinoma (6-7, 87, 89). According to Espanol et al., the expression of muscarinic acetylcholine receptor in murine mammary adenocarcinoma cells may modulate their aggressiveness (90). However, published data on the role of vagal innervation in the development of rat fibrosarcoma are inconclusive, and we still do not know if formation of metastases could be controlled with an inhibitor of the PNS signaling pathway (91). Nevertheless, Magnon et al. confirmed that autonomic nervous system plays a role in the development and spread of human prostate cancer (7). Tumor cholinergic signals are mediated by stromal chrm1 expression. Cholinergic fibres of PNS play role in tumor cell invasion, migration and distant metastases (7).

PNS plays an important role in the inflammatory reflex. Inflammation is a driving force for the dissemination and thus, inflammatory pathways and their effector cells, both controlled by the nervous system, contribute substantially to cancer spread. Inflammatory signals generated within the bowel may significantly alter peripheral neuronal signaling, which results in both peripheral and central sensitization, a phenomenon that is reflected by an enhanced afferent neuronal activation (92). As shown recently, the brain not only can "detect" peripheral inflammation via afferent vagal fibers, but may also attenuate innate immune activation due to an integrated neural response involving massive activation of vagal efferent fibers. This efferent arm of the inflammatory reflex is referred to as the "cholinergic anti-inflammatory pathway". The inflammatory reflex is a key

system involved in maintenance of homeostasis. Vagus nerve, the arc of the reflex and neural-related factors, such as netrin-1 and neuropeptides, all participate in the inflammation control (93, 94). Moreover, activation of vagus nerve exerts an effect on the production of cytokines by leukocytes. The electrostimulation of the vagus results in a decrease in cytokine production in the spleen, and an opposite effect has been observed after surgical ablation of this nerve (93). The cholinergic anti-inflammatory pathway requires signaling from the nicotinic acetylcholine receptor subunit  $\alpha 7$ , which inhibits the splenic nerve to suppress cytokine release by splenic macrophages (95).

Vagus nerve can influence immune cell function in the spleen through preganglionic and postganglionic system of neurons. The preganglionic system originates from the dorsal motor nucleus of the vagus, and the postganglionic system, from the ganglia of the celiac superior mesenteric plexus (94).

### **Role of non-cholinergic and non-adrenergic projections in carcinogenesis**

Dopamine produced in the brain is a catecholamine. However, unlike for norepinephrine and epinephrine, only few reports describe the role of dopamine in tumor cell migration and metastasis (52, 96-97). Dopamine participates in regulating gene expression, such as induced tumor cell migration. It acts through dopamine receptors (DRs) activation which are members of seven transmembrane domain trimeric guanosine 5'-triphosphate (GTP)-binding protein-coupled receptor family (97). DRD2/DARPP-32 expression is associated with tumor progression, therefore DRD2/DARPP-32 expressions can be a predictive factor (97).

Chemokines bind to receptors from the G-protein-coupled receptor (GPCR) family to regulate tumor-associated angiogenesis and tumor-specific immune response of the host, and to stimulate tumor cell proliferation in an autocrine manner (98). These findings may be the potential target of the therapy for cancer patients.

Gamma-aminobutyric acid (GABA) is an inhibitory neurotransmitter of the brain and an inhibitor of

glucagon secretion in the pancreatic  $\alpha$  cells. Involvement of GABA receptors was demonstrated to block the migratory effect of norepinephrine in mammary, pancreatic and colon carcinoma (52, 99-100).

Substance P acts as a neurotransmitter, neuro-modulator and inflammatory mediator. However, this molecule can be also involved in the development of bone marrow metastases of breast cancer and neuroblastoma (101). Moreover, substance P was shown to stimulate growth of basal-like human breast carcinoma cell line, MDA-MB-468 (52).

Taken altogether, those findings suggest that GABA-receptor agonists and the blockade of substance P receptor (NK-1) may constitute novel strategies in anticancer therapy.

### **ANS and Heart Rate Variability**

In a clinical setting, the autonomic activity can be assessed based on Heart Rate Variability (HRV) analysis. Monitoring of HRV is a non-invasive method to examine autonomic innervation of the heart and the vegetative modulation of the sinus node. The time- and frequency-domain analysis of the HRV provides an information on the autonomic balance. Furthermore, HRV is suitable for the quantification of sympathetic and parasympathetic tone (102). HRV analysis is a routine, widely-available method to evaluate vagal activity based on several indices, such as standard deviation of all normal beat to beat intervals (SDNN), root mean square successive difference between adjacent normal beat to beat intervals (RMSSD), the number of pairs of adjacent NN intervals differing by more than 50 ms in the entire recording divided by the total number of all NN intervals (pNN50), and high frequency (0.15-0.4 Hz) component (HF). Low values of the HRV analysis parameters may reflect sympathetic predominance or a decrease in parasympathetic input (102).

Vagal tone, expressed by the HRV, may constitute a new independent prognostic factor in patients with some solid tumors (103-105). Enhanced vagal activity may slow down tumorigenesis and produce a protective effect in patients with advanced or/and metastatic cancers (106). Several studies demonstrated a positive

association between high-frequency HRV component and overall survival in cancer patients; this relationship was independent of potential confounders, such as anticancer treatment, patient age and sex (106).

Previous studies demonstrated that prognosis in cancer is inter alia determined by the parasympathetic activity. De Couck et al. evaluated ANS in subjects with colorectal, pancreatic, prostate, lung and ovarian malignancies; the study demonstrated that cancer patients presented with significantly lower values of HRV parameters (SDNN and RMSSD) than healthy controls (103). Furthermore, the values of those HRV indices turned out to be significantly lower in patients with advanced cancers than in individuals with early malignancies. Other factors, such as age and sex, did not exert a significant effect on HRV analysis parameters in the study groups (103).

Another study revealed that vagal activity may be a prognostic factor in cancer patients. Patients diagnosed with colorectal and prostate cancers presented with higher levels of carcinoembryonic antigen (CEA) at 12 months and prostate-specific antigen (PSA) at 6 months, respectively, whenever their values of HRV indices were low (104). Furthermore, a positive correlation was found between vagal activity and overall survival in patients with advanced pancreatic cancer. According to Mouton et al., vagal activity may play a role in cancer recurrence (105). As widely known, the level of CEA, a cancer marker, correlates with the risk of relapse in most colorectal cancer patients. However, the study revealed that 12 months after anticancer treatment, patients who had increased vagal tone at the baseline (SDNN > 20 ms) presented with lower levels of CEA than individuals with lesser HRV (SDNN < 20 ms). Moreover, higher baseline HRV turned out to be associated with longer overall survival, also after adjustment for age and anticancer treatment (106). Moreover, high-frequency HRV component, a measure of parasympathetic overactivity, was shown to correlate positively with time to death in patients with terminal hepatocellular carcinoma and non-lung cancer (107, 108).

These findings may explain why prognosis in some cancer patients is worse than in the others; this phenomenon may be related to autonomic dysregulation of cardiac function. A dysfunction of ANS in cancer

patients, especially sympathovagal disturbances, may contribute to changes in cardiovascular system regulation. Lower values of HRV indices (markers of vagal activity) are associated with increased risk for life-threatening arrhythmias, which is reflected by generally worse prognosis.

### **ANS and cancer-induced fatigue**

Cancer-related fatigue has been long time ago recognized as a multi-factorial problem present in cancer patients. The issue has been inter alia addressed in a trial including breast cancer survivors who had completed anti-cancer treatment within past two years. Compared to their less-fatigued counterparts, the subjects who reported more fatigue had lower parameters of HRV analysis (only RMSSD was evaluated) and presented with significantly higher norepinephrine levels prior to and after the exposure to the study stressor. Those findings point to a potential link between a decrease in parasympathetic activity and greater fatigue. The level of fatigue was associated neither with the type of anticancer treatment nor with the type and stage of the malignancy. However, female patients who reported more fatigue were significantly older and showed age-specific changes in HRV; this implies that cancer-related fatigue may increase with age (109). A link between a decrease in selected HRV indices (indicators of parasympathetic activity) and greater fatigue in breast cancer patients has been also reported by Crosswell et al. (110). In the study conducted by those authors, lower values of two HRV indices, RMSSD and HF, correlated not only with greater fatigue, but also with older age, higher values of body mass index (BMI) and higher concentrations of IL-6 and C-reactive protein (CRP). However, the relationship between HRV and fatigue did not seem to be modulated by inflammatory mediators (110).

### **Importance of Autonomic Malfunction for Clinical Practice and .....**

Epinephrine and norepinephrine act via  $\beta$ -adrenergic receptor signaling pathway. Both preclin-

ical and in vitro studies demonstrated that adrenergic activation modulates apoptosis, promotes angiogenesis and other cancer hallmarks; all these effects can be abrogated by  $\beta$ -blockers. Therefore, treatment with  $\beta$ -adrenergic antagonists (e.g.  $\beta$ -blockers used widely in cardiology) may be a new therapeutic strategy to control of tumor progression (4, 111-112). There are three categories of beta-blockers. First generation  $\beta$ -blockers, i.e. non-selective  $\beta$ -blockers, such as propranolol, are antagonists of both  $\beta_1$  and  $\beta_2$  adrenergic receptors. Second generation  $\beta$ -blockers (e.g. atenolol) show higher affinity for  $\beta_1$ - than for  $\beta_2$ -adrenergic receptors. Third generation  $\beta$ -blockers (e.g. nebivolol) exert vasodilatory effects (113). Previous studies examined the effects of  $\beta$ -blockers in some malignancies, such as breast cancer, melanoma, pancreatic cancer and oral squamous cell cancer (62, 85, 114, 115). Pharmacological inhibition of  $\beta$ -adrenergic receptors resulted in downregulation of VEGF, and promoted apoptosis in cancer cells (54, 116). This implies that  $\beta$ -adrenergic receptor pathways may constitute a target for anticancer therapies; this application of  $\beta$ -blockers has been already tested in patients with various malignancies. In a retrospective study of patients with triple negative cancer, conducted by Melhem-Bertrandt et al., administration of  $\beta$ -blockers concomitant to neoadjuvant chemotherapy was associated with prolonged relapse-free survival (RFS), but not overall survival (OS) (117). In another study, women taking propranolol or atenolol have been diagnosed with less advanced breast cancers than those who did not receive  $\beta$ -blockers (118). This observation is consistent with the results of a previous study in which  $\beta_2$ -adrenergic signaling pathway blockers inhibited progression of breast cancer. In the latter study, women who have been treated with  $\beta$ -blockers prior to breast cancer diagnosis, had a 57% lesser risk of tumor spread and a 71% lower cancer-specific 10-year mortality rate than other study subjects (119). Administration of  $\beta$ -blockers during the course of platinum-based chemotherapy was also identified as an independent positive prognostic factor in epithelial ovarian cancer patients after a cytoreductive surgery. Women from  $\beta$ -blocker group had longer median progression-free survival and median overall cancer-specific survival than other study subjects, in 27 vs. 17 months and 56 vs. 48 months, respectively

(120). Also in a retrospective cohort study of patients with non-small-cell lung cancer, administration of  $\beta$ -blockers turned out to be associated with improved metastasis-free survival, disease-free survival and overall survival (121).

Another study conducted in Denmark demonstrated that administration of  $\beta$ -blockers was associated with a lower mortality risk in melanoma patients (122). However, another study did not document a significant effect of  $\beta$ -blockers, propranolol or atenolol, on colorectal cancer-specific mortality (123). Also, in a Norwegian cohort study of prostate cancer patients, administration of  $\beta$ -blockers was not associated with cancer-specific and overall mortality, PSA level, Gleason score and tumor stage at diagnosis. However, the same study demonstrated a significant decrease in cancer-specific mortality in a subgroup of men who received a  $\beta$ -blocker together with androgen deprivation therapy (ADT) (124). Administration of  $\beta$ -blockers did not contribute to significant differences in median progression-free survival and overall survival in patients with recurrent platinum-sensitive ovarian cancer, also after adjustment for age, platinum-based therapy-free interval, study treatment and performance status (125).

Despite numerous preclinical and clinical studies, we still need more evidence to confirm those findings. The results of some previous studies are inconclusive. Furthermore, we still do not know the optimal timing and duration of  $\beta$ -blocker treatment. At least 36 clinical studies analyzing the applicability of  $\beta$ -blockers in anticancer treatment are either currently ongoing or have just been completed, and we still wait for publication of their results.

#### .... and Prevention

Functioning of the neurotransmitter pathways may be affected by many extrinsic factors, such as lifestyle, exercise, physical activity, diet, concomitant diseases (e.g. cardiovascular disorders, diabetes mellitus), stress, unfavorable psychosocial work conditions and pharmacotherapy (126-128). Nicotinic acetylcholine receptors regulate the stimulation and inhibition of neurotransmitters and consequently, control the syn-

thesis and release of growth factors, angiogenic factors and neurotrophic factors in cancer cells, tumor microenvironment and metastatic foci. Some lifestyle-related factors (e.g. smoking) may upregulate nicotinic acetylcholine receptors and thus, contribute to stimulation of cancer cells (90). Therefore, healthy lifestyle with adequate level of physical activity seems to be crucial not only for cardiovascular prevention but also for the reduction of cancer risk.

## Conclusions

Knowledge of interactions between cancer cells and ANS seems to be a key for the development of novel anticancer therapies. However, to develop such tailored anticancer treatments, we need more information about the role of sympathetic and parasympathetic pathways in various malignancies.

## References

1. Wolter JK, Wolter NE, Blanch A, et al. Anti-tumor activity of the beta-adrenergic receptor antagonist propranolol in neuroblastoma. *Oncotarget* 2014; 5: 161-172.
2. Reeder A, Attar M, Nazario L, et al. Stress hormones reduce the efficacy of paclitaxel in triple negative breast cancer through induction of DNA damage. *Br J Cancer* 2015; 112: 1461-1470.
3. Cole SW, Sood AK. Molecular pathways: beta-adrenergic signaling in cancer. *Plos ONE* 2012; 18: 1201-1206.
4. Entschladen F, Drell TL 4th, Lang K, Joseph J, Zaenker KS. Tumour-cell migration, invasion, and metastasis: navigation by neurotransmitters. *Lancet Oncol* 2004; 5: 254-258.
5. Wong SY, Hynes RO. Lymphatic or hematogenous dissemination: how does a metastatic tumor cell decide? *Cell Cycle* 2006; 5: 812-817.
6. Fernandez EV, Price DK, Figg WD. Prostate cancer progression attributed to autonomic nerve development. Potential for therapeutic prevention of localized and metastatic disease. *Cancer Biol Ther* 2013; 14: 1005-1006.
7. Magnon C, Hall SJ, Lin J, et al. Autonomic nerve development contributes to prostate cancer progression. *Science* 2013; 341: 1236361.
8. Voss MJ, Entschladen F. Tumor interactions with soluble factors and the nervous system. *Cell Commun Signal* 2010; 8. doi: 10.1186/1478-811X-8-21.
9. Magnon C. Role of the autonomic nervous system in tumorigenesis and metastasis. *Mol Cell Oncol* 2015; 2:e975643.
10. Ayala GE, Dai H, Powell M, et al. Cancer-related axonogenesis and neurogenesis in prostate cancer. *Clin Cancer Res* 2008; 14: 7593-603.
11. Sastry KS, Karpova Y, Prokopovich S, et al. Epinephrine protects cancer cells from apoptosis via activation of cAMP-dependent protein kinase and BAD phosphorylation. *J Biol Chem* 2007; 282: 14094-14100.
12. Cole SW, Nagaraja AS, Lutfgendorf SK, Green PA, Sood AK. Sympathetic nervous system regulation of the tumour microenvironment. *Nat Rev Cancer* 2015; 15: 563-572.
13. Bacac M, Stamenkovic I. Metastatic cancer cell. *Annu Rev Pathol* 2008; 3: 221-247.
14. Raquel del Toro, Simón Méndez-Ferrer. Autonomic regulation of hematopoiesis and cancer. *Haematologica* 2013; 98: 1663-1666.
15. Li S, Sun Y, Gao D. Role of the nervous system in cancer metastasis. *Oncol Lett* 2013; 5: 1101-1111.
16. Liebig C, Ayala G, Wilks JA, Berger DH, Albo D. Perineural invasion in cancer: a review of the literature. *Cancer* 2009; 115: 3379-3391.
17. Goldberg JL. How does an axon grow? *Genes Dev* 2003; 17: 941-958.
18. Chilton JK. Molecular mechanisms of axon guidance. *Dev Biol* 2006; 292: 13-24.
19. Chen C, Li M, Chai H, Yang H, Fisher WE, Yao Q. Roles of neuropilins in neuronal development, angiogenesis, and cancers. *World J Surg* 2005; 29: 271-275.
20. Kruger RP, Aurandt J, Guan KL. Semaphorins command cells to move. *Nat Rev Mol Cell Biol* 2005; 6: 789-800.
21. Entschladen F, Palm D, Drell TL 4th, Lang K, Zaenker KS. Connecting a tumor to the environment. *Curr Pharm Des* 2007; 13: 3440-3444.
22. Chedotal A. Chemotropic axon guidance molecules in tumorigenesis. *Prog Exp Tumor Res* 2007; 39: 78-90.
23. Palm D, Entschladen F. Neoneurogenesis and the neuro-neoplastic synapse. *Prog Exp Tumor Res* 2007; 39: 91-98.
24. Calza L, Giardino L, Giuliani A, Aloe L, Levi-Montalcini R. Nerve growth factor control of neuronal expression of angiogenic and vasoactive factors. *Proc Natl Acad Sci USA* 2001; 98: 4160-4165.
25. Kruttgen A, Schneider I, Weis J. The dark side of the NGF family: neurotrophins in neoplasias. *Brain Pathol* 2006; 16: 304-310.
26. Jimenez-Andrade JM, Bloom AP, Stake JI, et al. Pathological sprouting of adult nociceptors in chronic prostate cancer-induced bone pain. *J Neurosci* 2010; 30: 14649-14656.
27. Marko SB, Damon DH. VEGF promotes vascular sympathetic innervation. *Am J Physiol Heart Circ Physiol* 2008; 294: H2646-H2652.
28. Lazarovici P, Marcinkiewicz C, Lelkes PI. Cross talk between the cardiovascular and nervous systems: neurotrophic effects of vascular endothelial growth factor (VEGF) and angiogenic effects of nerve growth factor (NGF)-implications in drug development. *Curr Pharm Des* 2006; 12: 2609-2622.

29. Yasuhara T, Shingo T, Date I. The potential role of vascular endothelial growth factor in the central nervous system. *Rev Neurosci* 2004; 15: 293-307.
30. Sondell M, Lundborg G, Kanje M. Vascular endothelial growth factor has neurotrophic activity and stimulates axonal outgrowth, enhancing cell survival and Schwann cell proliferation in the peripheral nervous system. *J Neurosci* 1999; 19: 5731-5740.
31. Ayala GE, Dai H, Ittmann M, et al. Growth and survival mechanisms associated with perineural invasion in prostate cancer. *Cancer Res* 2004; 64: 6082-90.
32. Dai H, Li R, Wheeler T, et al. Enhanced survival in perineural invasion of pancreatic cancer: an in vitro approach. *Human Pathol* 2007; 38: 299-307.
33. Guillemot F, Zimmer C. From cradle to grave: the multiple roles of fibroblast growth factors in neural development. *Neuron* 2011; 71: 574-588.
34. Dobrenis K, Gauthier LR, Barroca V, Magnon C. Granulocyte colony-stimulating factor off-target effect on nerve outgrowth promotes prostate cancer development. *Int J Cancer* 2015; 136: 982-988.
35. Zaenker KS. The neuro-neoplastic synapse: does it exist? *Prog Exp Tumor Res* 2007; 39: 154-61.
36. Olofsson PS, Rosas-Ballina M, Levine YA, Tracey KJ. Rethinking inflammation: neural circuits in the regulation of immunity. *Immunol Rev* 2012; 248: 188-204.
37. Kenney MJ, Ganta CK. Autonomic nervous system and immune system interactions. *Compr Physiol* 2014; 4: 1177-1200.
38. Tan W, Zhang W, Strasner A, Grivennikov S, Cheng JQ, Hoffman RM, Karin M. Tumour-infiltrating regulatory T cells stimulate mammary cancer metastasis through RANKL-RANK signalling. *Nature* 2011; 470: 548-553.
39. Reiche EM, Nunes SO, Morimoto HK. Stress, depression, the immune system, and cancer. *Lancet Oncol* 2004; 5: 617-625.
40. Elenkov IJ, Wilder RL, Chrousos GP, Vizi ES. The sympathetic nerve--an integrative interface between two super-systems: the brain and the immune system. *Pharmacol Rev* 2000; 52: 595-638.
41. Hasko G, Szabo C. Regulation of cytokine and chemokine production by transmitters and co-transmitters of the autonomic nervous system. *Biochem Pharmacol* 1998; 56: 1079-1087.
42. Hanoun M, Maryanovich M, Arnal-Estape A, Frenette PS. Neural regulation of hematopoiesis, inflammation, and cancer. *Neuron* 2015; 86: 360-373.
43. Kawashima K, Fujii T. The lymphocytic cholinergic system and its contribution to the regulation of immune activity. *Life Sci* 2013; 74: 675-696.
44. Kafetzopoulou LE, Boocock DJ, Dhondalay GK, Powe DG, Ball GR. Biomarker identification in breast cancer: Beta-adrenergic receptor signaling and pathways to therapeutic response. *Comput Struct Biotechnol J* 2013; 6: e201303003. doi: 10.5936/csbj.201303003, Ecollection.
45. Armaiz-Pena GN, Allen JK, Cruz A, et al. Src activation by  $\beta$ -adrenoreceptors is a key switch for tumour metastasis. *Nat Commun* 2013; 4: 1403.
46. Gu L, Lau SK, Loera S, Somlo G, Kane SE. Protein kinase A activation confers resistance to trastuzumab in human breast cancer cell lines. *Clin Cancer Res* 2009; 15: 7196-7206.
47. Shi M, Liu D, Duan H, et al. The  $\beta$ 2-adrenergic receptor and Her2 comprise a positive feedback loop in human breast cancer cells. *Breast Cancer Res Treat* 2011; 125: 351-362.
48. Hara MR, Kovacs JJ, Whalen EJ, et al. A stress response pathway regulates DNA damage through  $\beta$ 2-adrenoreceptors and  $\beta$ -arrestin-1. *Nature* 2011; 477: 349-353.
49. Hara MR, Sachs BD, Caron MG, Lefkowitz RJ. Pharmacological blockade of a  $\beta(2)$ AR- $\beta$ -arrestin-1 signaling cascade prevents the accumulation of DNA damage in a behavioral stress model. *Cell Cycle* 2013; 12: 219-224.
50. Daly CJ, McGrath JC. Previously unsuspected widespread cellular and tissue distribution of  $\beta$ -adrenoreceptors and its relevance to drug action. *Trends Pharmacol Sci* 2011; 32: 219-226.
51. Palm D, Lang K, Niggemann B, et al. The norepinephrine-driven metastasis development of PC-3 human prostate cancer cells in BALB/c nude mice is inhibited by beta-blockers. *Int J Cancer* 2006; 118: 2744-2749.
52. Drell TL 4th, Joseph J, Lang K, Niggemann B, Zaenker KS, Entschladen F. Effects of neurotransmitters on the chemokinesis and chemotaxis of MDA-MB-468 human breast carcinoma cells. *Breast Cancer Res Treat* 2003; 80: 63-70.
53. Lang K, Drell TL 4th, Lindecke A, et al. Induction of a metastatogenic tumor cell type by neurotransmitters and its pharmacological inhibition by established drugs. *Int J Cancer* 2004; 112: 231-238.
54. Thaker PH, Han LY, Kamat AA, et al. Chronic stress promotes tumor growth and angiogenesis in a mouse model of ovarian carcinoma. *Nat Med* 2006; 12: 939-944.
55. Chakroborty D, Sarkar C, Basu B, Dasgupta PS, Basu S. Catecholamines regulate tumor angiogenesis. *Cancer Res* 2009; 69: 3727-3730.
56. Yang EV, Sood AK, Chen M, et al. Norepinephrine up-regulates the expression of vascular endothelial growth factor, matrix metalloproteinase (MMP)-2, and MMP-9 in nasopharyngeal carcinoma tumor cells. *Cancer Res* 2006; 66: 10357-10364.
57. Sood AK, Bhatti R, Kamat AA, et al. Stress hormone-mediated invasion of ovarian cancer cells. *Clin Cancer Res* 2006; 12: 369-375.
58. Landen CN jr, Lin YG, Armaiz Pena GN, et al. Neuroendocrine modulation of signal transducer and activator of transcription-3 in ovarian cancer. *Cancer Res* 2007; 67: 10389-10396.
59. Draoui A, Yandewalle B, Hornez L, Revillion F, Lefebvre J. Beta-adrenergic receptors in human breast cancer: identification, characterization and correlation with progesterone and estradiol receptors. *Anticancer Res* 1991; 11: 677-680.
60. Slotkin TA, Zhang J, Dancel R, Garcia SJ, Willis C, Seidler

- FJ. Beta-adrenoceptor signaling and its control of cell replication in MDA-MB-231 human breast cancer cells. *Breast Cancer Res Treat* 2000; 60: 153-166.
61. Sloan E, Priceman SJ, Cox BF, et al. The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res* 2010; 70: 7042-7052.
62. Barron TI, Sharp L, Visvanathan K. Beta-adrenergic blocking drugs in breast cancer: a perspective review. *Ther Adv Med Oncol* 2012; 4: 113-125.
63. Bauman J, McVary K. Autonomic nerve development contributes to prostate cancer progression. *Asian J Androl* 2013; 15: 713-714.
64. Shi M, Liu D, Yang Z, Guo N. Central and peripheral nervous systems: master controllers in cancer metastasis. *Cancer Metastasis Rev* 2013; 32: 603-621.
65. Horvathova L, Mravec B. Effect of the autonomic nervous system on cancer progression depends on the type of tumor: solid are more affected than ascitic tumors. *Endocrin Regul* 2016; 50: 215-224.
66. Dolle L, El Yazidi-Belkoura I, Adriaenssens E, Nurcombe V, Hondermarck H. Nerve growth factor overexpression and autocrine loop in breast cancer cells. *Oncogene* 2003; 22: 5592-5601.
67. Hondermarck H. Neurotrophins and their receptors in breast cancer. *Cytokine Growth Factor Rev* 2012; 23: 357-365.
68. Larrivee B, Freitas C, Suchting S, Brunet I, Eichmann A. Guidance of vascular development: lessons from the nervous system. *Circ Res* 2009; 104: 428-441.
69. Mehlan P, Delloye-Bourgeois C, Chedotal A. Novel roles for Slits and netrins: axon guidance cues as anticancer targets? *Nat Rev Cancer* 2011; 11: 188-197.
70. Neufeld G, Kessler O. The semaphorins: versatile regulators of tumour progression and tumour angiogenesis. *Nat Rev Cancer* 2008; 8: 632-645.
71. Potiron V, Roche J, Drabkin HA. Semaphorins and their receptors in lung cancer. *Cancer Lett* 2009; 273: 1-14.
72. Castro-Rivera E, Ran S, Brekken RA, Minna JD. Semaphorin 3B inhibits the phosphatidylinositol 3-kinase/Akt pathway through neuropilin-1 in lung and breast cancer cells. *Cancer Res* 2008; 68: 8295-8303.
73. Nakada M, Hayashi Y, Hamada J. Role of Eph/ephrin tyrosine kinase in malignant glioma. *Neuro Oncol* 2011; 13: 1163-1170.
74. Barquilla A, Pasquale EB. Eph receptors and ephrins: therapeutic opportunities. *Annu Rev Pharmacol Toxicol* 2015; 55: 465-487.
75. Eichmann A, Thomas JL. Molecular parallels between neural and vascular development. *Cold Spring Harb Perspect Med* 2013; 3: a006551. doi: 10.1101/cshperspect.a006551.
76. Damon DH, Teriele JA, Marko SB. Vascular-derived artemin: a determinant of vascular sympathetic innervation? *Am J Physiol Heart Circ Physiol* 2007; 293: H266-H273.
77. Makita T, Sucov HM, Garipey CE, Yanagisawa M, Ginty DD. Endothelins are vascular-derived axonal guidance cues for developing sympathetic neurons. *Nature* 2008; 452: 759-763.
78. Luo MY, Yang BL, Ye F, et al. Collateral vessel growth induced by femoral artery ligation is impaired by denervation. *Mol Cell Biochem* 2011; 354: 219-229.
79. Chalothorn D, Zhang H, Clayton JA, Thomas SA, Faber JE. Catecholamines augment collateral vessel growth and angiogenesis in hindlimb ischemia. *Am J Physiol Heart Circ Physiol* 2005; 289: 947-959.
80. Cohen MJ, Sahnkar R, Stevenson J, Fernandez R, Gamelli RL, Jones SB. Bone marrow norepinephrine mediates development of functionally different macrophages after thermal injury and sepsis. *Ann Surg* 2004; 240: 132-141.
81. Engler H, Bailey MT, Engler A, Sheridan JF. Effects of repeated social stress on leukocyte distribution in bone marrow, peripheral blood and spleen. *Neuroimmunol* 2004; 148: 106-115.
82. Laakko T, Fraker P. Rapid changes in the lymphopoietic and granulopoietic compartments of the marrow caused by stress levels of corticosterone. *Immunology* 2002; 105:111-119.
83. Xu Z, Shioda S, Masahisa J, et al. Role of the autonomic nervous system in the tumor micro-environment and its therapeutic potential. *Curr Pharm Des* 2017; 23: 1687-1692.
84. Madden KS, Szpunar MJ, Brown EB.  $\beta$ -Adrenergic receptors ( $\beta$ -AR) regulate VEGF and IL-6 production by divergent pathways in high  $\beta$ -AR-expressing breast cancer cell lines. *Breast cancer Res Treat* 2011; 130: 747-758.
85. Stock AM, Powe DG, Hahn SA, et al. Norepinephrine inhibits the migratory activity of pancreatic cancer cells. *Exp Cell Res* 2013; 319: 1744-1758.
86. Colucci R, Moretti S. The role of stress and beta-adrenergic system in melanoma: current knowledge and possible therapeutic options. *J Cancer Res Clin Oncol* 2016; 142: 1021-1029.
87. Ventura S, Evans BA. Does the autonomic nervous system contribute to the initiation and progression of prostate cancer? *Asian J Androl* 2013; 15: 715-716.
88. Obeid EI, Conzen SD. The role of adrenergic signaling in breast cancer biology. *Cancer Biomark* 2013; 13:161-169.
89. Isaacs JT. Cancer. Prostate cancer takes nerves. *Science* 2013; 341: 134-135.
90. Espanol A, Eijan AM, Mazzoni E, et al. Nitric oxide synthase, arginase and cyclooxygenase are involved in muscarinic receptor activation in different murine mammary adenocarcinoma cell lines. *Int J Mol Med* 2002; 9: 651-657.
91. Mikova L, Horvathova L, Ondicova K, et al. Ambiguous effect of signals transmitted by the vagus nerve on fibrosarcoma incidence and survival of tumor-bearing rats. *Neurosci Lett* 2015; 593: 90-94.
92. Huston JM. The vagus nerve and the inflammatory reflex: wandering on a new treatment paradigm for systemic inflammation and sepsis. *Surg Infect* 2012; 13: 187-193.
93. Sundman E, Olofsson PS. Neural control of the immune system. *Adv Physiol Educ* 2014; 38: 135-139.

94. Rosas-Ballina M, Ochani M, Parrish WR, et al. Splenic nerve is required for cholinergic antiinflammatory pathway control of TNF in endotoxemia. *Proc Natl Acad Sci USA* 2008; 105: 11008-11013.
95. Huston JM, Ochani M, Rosas-Ballina M, et al. Splenectomy inactivates the cholinergic antiinflammatory pathway during lethal endotoxemia and polymicrobial sepsis. *J Exp Med* 2006; 203: 1623-1628.
96. Bodei L, Hofland LJ, Ferone D, Mooy CM, Kros JM, Paridaens DA, et al. In vivo and in vitro detection of dopamine d2 receptors in uveal melanomas. *Cancer Biother Radiopharm* 2003; 18: 895-902.
97. Li L, Miyamoto M, Ebihara Y, Mega S, Takahashi R, Hase R, et al. DRD2/DARPP-32 expression correlates with lymph node metastasis and tumor progression in patients with esophageal squamous cell carcinoma. *Worlds J Surg* 2006; 30:1672-1679; discussion 1680-1681.
98. Kruizinga RC, Bestebroer J, Berghuis P, de Haas CJ, Links TP, de Vries EG, et al. Role of chemokines and their receptors in cancer. *Curr Pharm Des* 2009; 15:3396-3416.
99. Joseph J, Niggemann B, Zaenker KS, Entschladen F. The neurotransmitter gamma-aminobutyric acid is an inhibitory regulator for the migration of SW 480 colon carcinoma cells. *Cancer Res* 2002; 62: 6467-6469.
100. Schuller HM, Al-Wadei HA, Majidi M. GABA B receptor is a novel drug target for pancreatic cancer. *Cancer* 2008; 112: 767-778.
101. Nowicki M, Ostalska-Nowicka D, Kondraciuk B, Miskowiak B. The significance of substance P in physiological and malignant haematopoiesis. *J Clin Pathol* 2007; 60: 749-755.
102. Task Force of the European Society of Cardiology and The North American Society of Pacing and Electrophysiology: Heart rate variability. Standards of measurement, physiological interpretation and clinical use. *Circulation* 1996; 93: 1043-1065.
103. De Couck M, Gidron Y. Norms of vagal nerve activity, indexed by Heart Rate Variability, in cancer patients. *Cancer Epidemiol* 2013; 37: 737-741.
104. Gidron Y, De Couck M, De Greve J. If you have an active vagus nerve, cancer stage may no longer be important. *J Biol Regul Homeost Agents* 2014; 28: 195-201.
105. Mouton C, Ronson A, Razavi D, Delhaye F, Kupper N, Paesmans M, Gidron Y. The relationship between heart rate variability and time-course of carcinoembryonic antigen in colorectal cancer. *Auton Neurosc* 2012, 166: 96-99.
106. De Couck M, Marechal R, Moorthamers S, Van Laethem JL, Gidron Y. Vagal nerve activity predicts overall survival in metastatic pancreatic cancer, mediated by inflammation. *Cancer Epidemiol* 2014; 40: 47-51.
107. Fadul N, Strasser F, Palmer JL, Yusuf SW, Guo Y, Li Z, et al. The association between autonomic dysfunction and survival in male patients with advanced cancer: a preliminary report. *J Pain Symptom Manage* 2010; 39: 283-290.
108. Chiang JK, Kuo TB, Fu CH, Koo M. Predicting 7-day survival using heart rate variability in hospice patients with non-lung cancers. *PloS One* 2013; 8: e69482.
109. Fagundes CP, Murray DM, Hwang BS, et al. Sympathetic and parasympathetic activity in cancer-related fatigue: more evidence for a physiological substrate in cancer survivors. *Psychoneuroendocrinology* 2011; 36: 1137-1147.
110. Crosswell AD, Lockwood KG, Ganz PA, Bower JE. Low heart rate variability and cancer-related fatigue in breast cancer survivors. *Psychoneuroendocrinology* 2014; 45: 58-66.
111. Yang EV, Eubank TD. The impact of adrenergic signaling in skin cancer progression: possible repurposing of  $\beta$ -blockers for treatment of skin cancer. *Cancer Biomark* 2013; 13: 155-160.
112. Fitzgerald PJ. Beta blockers, norepinephrine and cancer: an epidemiological viewpoint. *Clin Epidemiol* 2012; 4: 151-156.
113. Nagaraja AS, Sadaoui NC, Lutgendorf SK, Ramondetta LH, Sood AK.  $\beta$ -blockers: a new role in cancer chemotherapy? *Expert Opin Investig Drugs* 2013; 22: 1359-1363.
114. De Giorgi V, Grazzini M, Gandini S, et al.  $\beta$ -adrenergic blocking drugs and melanoma: current state of the art. *Expert Rev Anticancer Ther* 2012; 12: 1461-1467.
115. Shang ZJ, Liu K, Liang DF. Expression of beta 2-adrenergic receptor in oral squamous cell carcinoma. *J Oral Pathol Med* 2009; 38: 371-376.
116. Hassan S, Karpova Y, Baiz D, et al. Behavioral stress accelerates prostate cancer development in mice. *J Clin Invest* 2013; 123: 874-886.
117. Melhem-Bertrandt A, Chavez-MacGregor M, Lei X, et al. Beta-blocker use is associated with improved relapse-free survival in patients with triple-negative breast cancer. *J Clin Oncol* 2011; 29: 2645-2652.
118. Barron TI, Connolly RM, Sharp L, Bennett K, Visvanathan K. Beta blockers and breast cancer mortality: a population-based study. *J Clin Oncol* 2011; 29: 2635-2644.
119. Powe DG, Voss MJ, Zaenker KS, et al. Beta-blocker drug therapy reduces secondary cancer formation in breast cancer and improves cancer specific survival. *Oncotarget* 2010; 1: 628-638.
120. Diaz ES, Karlan BY, Li AJ. Impact of beta blockers on epithelial ovarian cancer survival. *Gynecol Oncol* 2012; 127: 375-378.
121. Wang HM, Liao ZX, Komaki R, et al. Improved survival outcomes with the incidental use of beta-blockers among patients with non-small-cell lung cancer treated with definitive radiation therapy. *Ann Oncol* 2013; 24: 1312-1319.
122. Lemeshow S, Sorensen HT, Phillips G, et al.  $\beta$ -Blockers and survival among Danish patients with malignant melanoma: a population-based cohort study. *Cancer Epidemiol Biomarkers Prev* 2011; 20: 2273-2279.
123. Hicks BM, Murray LJ, Powe DG, Hughes CM, Cardwell CR.  $\beta$ -Blocker usage and colorectal cancer mortality: a nested case-control study in the UK Clinical Practice Research Datalink cohort. *Ann Oncol* 2013; 24: 3100-3106.

124. Grytli HH, Fagerland MW, Fossa SD, Tasken KA, Haheim LL. Use of  $\beta$ -blockers is associated with prostate cancer-specific survival in prostate cancer patients on androgen deprivation therapy. *Prostate* 2013; 73: 250-260.
125. Heitz F, Du Bois A, Harter P, et al., AGO study group; NCIC-CTG study group; EORTC-GCG study group: Impact of beta blocker medication in patients with platinum sensitive recurrent ovarian cancer-a combined analysis of 2 prospective multicenter trials by the AGO Study Group, NCIC-CTG and EORTC-GCG. *Gynecol Oncol* 2013; 129: 463-466.
126. Schuller HM. Is cancer triggered by altered signalling of nicotinic acetylcholine receptors? *Nat Rev Cancer* 2009; 9: 195-205.
127. Pietila J, Helander E, Myllymaki T, Korhonen I, Jimison H, Pavel M. Exploratory analysis of associations between individual lifestyles and heart rate variability - based recovery during sleep. *Conf Proc IEEE Eng Med Biol Soc* 2015; 2015: 2339-2342.
128. Fohr T, Pietila J, Helander E, et al. Physical activity, body mass index and heart rate variability-based stress and recovery in 16 275 Finnish employees: a cross-sectional study. *BMC Public Health* 2016; 16: 701.

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